Module name	Microscopic techniques
Module code	B-BC.BE.223E
ISCED code	0511: Biology
Study cycle	
Semester	summer
Responsible for this module	Main teacher Bożena Pawlikowska-Pawlęga,
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	Department of Functional Anatomy and Cytobiology
Language of instruction	English
Website	https://www.umcs.pl/en/biology-and-
	biotechnology,9741.htm.
Prerequisites	basic knowledge of English, the passed courses from
	cell biology, chemistry and biochemistry
ECTS	3
ECTS points hour equivalents	30 Contact hours with the lecturer in the form of classes
	10 Student preparation for didactic classes
	20 Student preparation for credit
	15 Studying subject literature by a student
	Total number of ECTS points for the module -
Learning outcomes verification methods	mid-term tests (W1-W5, U1-U4, K1-K2)
	attendance at exercises
	K-K01: attendance at lectures
Course full description	1. Construction of electron microscopes and their
	functioning. Characteristics of SEM and TEM
	parameters.
	parameters.
	2. Visit in TEM ansd SEM labs. Microscopic observation
	of specimens from the selected cells, tissues, animal
	organs, viruses and bacteria.
	3. Construction and operation of confocal microscope.
	Advanced Techniques dedicated for confocal
	microscopy: FRET (Fluorescence / Forster resonance
	energy transfer), FRAP (fluorescence recovery after
	photobleaching), FLIP (Fluorescence loss in
	photobleaching), FLIM (fluorescence lifetime imaging microscopy).
	4. Visit in confocal microscopy lab of UMCS. Slides
	observation and saving images in confocal microscope.
	Convertion to three-dimensional image.

5. Test - electron microscopy: transmission, scanning
and confocal microscopy.
6. Construction and operation of a light microscope (dark and bright field of view, phase contrast and fluorescence).
7. Preparation of samples for electron microscopy - the whole procedure. Collecting material, fixation, dehydration, saturation and embedding with resin, cutting on ultrathin sections, contrasting . Physical and chemical fixation. Carriers for fixatives and the criteria for their selection .
8. Trimming and grids preparation for electron microscopy. Manual trimming and trimming on ultramicrotome.
9. Cutting of biological samplesCutting material into
silver sections. Straightening of sections with
chloroform. Collecting sections on copper grids.
10. Positive grid's contrasting. Positive contrasting -
factors relevant for contrasting. Contrasting
compounds: osmium tetroxide, uranyl acetate, lead
salts, phosphotungstic acid, potassium permanganate.
Negative contrasting. Contrasting compounds used in
contrasting: uranyl acetate, Reynolds reagent :lead
nitrate, sodium citrate.
11. Test- Construction and operation on light
microscope (bright field, dark field, phase contrast and
fluorescence; samples preparation for electron
microscopy - all stages of the procedure.
12. Slides observation with application of light
microscope.
Staining of human cheek epithelium with methylene
blue. HeLa cells from culture in vitro –semithin section
stained with toluidine blue. Cerebellar cortex cells,
hyaline cartilage of human trachea , squamous epithelial

	cells of the esophagus, ureter epithelium, seminiferous
	tubules of rat;
	13. Preparation of sample for SEM from cells in vitro.
	Coating of sample with usage of coat sputter and
	recording images in electron microscope (SEM).
	14. Slides observation in fluorescence microscopy.
	Identification of autophagic cells – acridine orange
	staining; Apototic and necrotic cells identification –
	staining with propidium iodide and Hoechst 33342.
	15. Final assessment.
Pibliography	1. Alberts B, BrayD, Johnson A, lewis J, Raff M, Roberts
Bibliography	K, Walter P. Podstawy biologii komórki. PWN 2005.
	2. Immunocytochemia. PWN 1999.
	3. Klyszejko-Stefanowicz L. Cytobiochemia. PWN 2002.
	4. Reid N. Ultramicrotomy [in:] Glanert AM. Practical
	methods in electron microscopy. Vol. 3,1975.
	5. J. Litwin, M. Gajda, WUJ 2011, Podstawy Technik
	Mikroskopowych.
	6. B. Wróbel, K. Zienkiewicz, D. Smoliński, J. Niedojadało, M. Świdziński, WUMK 2005, Podstawy Mikroskopii Elektronowej
Learning outcomes	KNOWLEDGE
	K1. The student interprets the images visible on the preparations, i.e. he recognizes autophagy, apoptosis,
	normal and changed structure of tumor and tumor cells
	K2. The student has in-depth knowledge of physics and
	chemistry to the extent necessary to understand the
	theoretical foundations of the microscopic techniques
	used, including electron, confocal, light and
	fluorescence microscopy, i.e. he knows the
	phenomenon of fluorescence, phosphorescence, elastic and non-elastic scattering of electrons, diffraction,
	phase shift, chemical fixation in connection with other
	fields of science especially with physics and chemistry
	K3. Understands the principles of selection of individual
	microscopic methods and techniques used in biological

	<ul> <li>sciences, i.e. for TEM ultrastructure research, for SEM</li> <li>surface topography, for life-long research - fluorescence</li> <li>and confocal microscopy</li> <li>K4. Knows the principles of safe and ergonomic work in</li> <li>the laboratory, especially during preparation</li> <li>procedures during fixation, trimming and work with</li> <li>normal and cancer cells from in vitro culture</li> </ul>
	SKILLS
	S1. The graduate is able to select and use appropriate microscopic techniques for specific research purposes and modify standard procedures to achieve specific goals
	S2. The graduate is able to cooperate with other people as part of team work (trimming, preparation of preparations) in performing various tasks in the field of microscopic techniques, i.e. fluorescence, SEM and TEM and light microscopy
	SOCIAL COMPETENCES
	S1. The graduate is ready to lead the group and take responsibility for it and make decisions during the implementation of practical parts of the entire cycle of exercises
	S2. The graduate is ready to recognize the importance of knowledge in the field of microscopic techniques, i.e. in scientific, medical and pharmacological research (FLIM, TEM, EDX), related sciences, i.e. physics and chemistry in solving cognitive and practical problems
Practice	
Teaching methods	explanation, explanation, observation, practical preparation of samples and elements of the procedure for electron and confocal microscopy; lecture